

## Mitral Valve Prolapse: Current Views and Challenges (Review)

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Mitral valve prolapse (MVP) is the most common valve abnormality. Many issues relating its diagnosis, epidemiology, prognosis, and genetics have lately been defined more precisely or revised.

The most principal changes in MVP diagnosis are connected with establishing a three-dimensional saddle-like shape of the mitral valve annulus, which made mandatory the assessment of the valve condition from the parasternal longitudinal position during ultrasound examination. Implementation of standard diagnostic criteria based on two-dimensional echocardiography, and making the results of the Framingham Heart Study public made it possible to overcome the contradictions relative to the prevalence of this pathology, which appeared to be lower than it had been considered earlier. Age, gender, and ethnic characteristics of MVP occurrence have been established. Notions not only about the incidence of mitral prolapse development but the severity of its sequelae were subjected to reassessment. If previously MVP was thought to be a disease with serious complications, findings of conducted epidemiological studies gave reasons to consider it as a benign pathology with a low probability of unfavorable consequences. Concurrently, factors of unfavorable prognosis were identified, and mitral regurgitation was recognized to be the main of them.

The results of molecular genetic investigations enriched essentially notion about MVP and improved its diagnosing. At present, this pathology is believed to be a result of multiple genetic disorders caused by identification of several genes linked with the onset of syndromic prolapse, and three loci for nonsyndromic one. Creation of large-scale registers of MVP patients and conduction of genome-wide studies will enable cardiologists to identify new genes related to the emergence of mitral prolapse and provide screening of asymptomatic patients. The leading role in various mechanisms of MVP pathogenesis is played by the impairment of regulation of transforming growth factor beta (TGF- $\beta$ ), understanding of pathogenetic role of which opens new perspectives of conservative treatment of this pathology with the application of antibodies neutralizing TGF- $\beta$ , and angiotensin II receptor blockers. Such medical approaches may be rather promising at the early stage of undiagnosed MVP phenotypes, and also serve as an alternative to surgical treatment of clinical complications in patients with a verified diagnosis.

**Key words:** mitral valve prolapse; mitral prolapse diagnosing; MVP epidemiology; prognosis in prolapses; molecular and genetic basics in MVP; mitral regurgitation; transforming growth factor beta.

Mitral valve prolapse (MVP) is the most common valve abnormality, which occurs in 2–3% of population [1–5]. This pathology is thought to be the leading cause of isolated mitral insufficiency demanding surgical intervention [3, 6–8]. Prolapse is known to be primary and secondary. The secondary (syndromic) MVP is the result of monogenic defects of connective tissue such as Marfan, Loeys–Dietz, Ehlers–Danlos syndromes, osteogenesis imperfecta, pseudoxanthoma elasticum, and recently described aneurysms-osteoarthritis syndrome [8–12]. Typical MVP is characterized by

myxomatous degeneration of mitral leaflets and their systolic displacement to the left atrium cavity [13, 14].

In recent years, many notions about diagnosis, epidemiology, prognosis, and genetics of MVP have been made more precise or revised, the most important directions of further investigations have been specified.

### Diagnosis of mitral valve prolapse

Clinical picture of MVP is very heterogeneous; mitral prolapse can be asymptomatic or have clinical

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manifestations [4, 6, 15, 16]. Physical examination supplemented by two-dimensional echocardiography remains a golden standard of MVP diagnosis [17–19].

A leading mechanism disclosing a diverse MVP semiotics is vegetative dysfunction, though the availability of asymptomatic patients does not permit an unambiguous definition of its pathogenetic role [20, 21]. Nevertheless, the majority of researchers [22–24] consider vegetative changes of homeostasis to be the obligate MVP manifestation. Quite a number of hypotheses have been suggested explaining presence of vegetative dysfunction in MVP, including congenital alterations of perineurium, a systemic defect of biological membranes, perinatal lesion of hypothalamic structures [20, 25, 26], and a version about a pathogenetic role of hypomagnesemia being lately actively discussed [27].

When two-dimensional echocardiography was first implemented into clinical practice, MVP was diagnosed in 5–15% and even in 35% of the examined patients [28, 29]. Such results were mainly connected with erroneous assumption that a mitral valve has a plane configuration. A series of ultrasound examinations [18, 30, 31] allowed the physicians to establish that a mitral valve annulus was saddle-shaped, and made the assessment of the valve condition from the parasternal longitudinal position obligatory [32, 33]. Modern medicine defines MVP as a systolic bulging of one or both mitral valve leaflets by no less than 2 mm beyond the mitral annulus plane with its obligatory long-axis registration [5].

Three-dimensional echocardiography improved essentially understanding of the mechanics of normal and pathologic mitral valves. This method is of value not only in MVP diagnosis but in determining the tactics of its surgical treatment and outcome assessment [18, 34–38].

### Prevalence of mitral valve prolapse

Notions about the MVP prevalence remained controversial till the announcement of the results of the community-based Framingham Heart Study [1, 39, 40], and transition to the unified criteria of ultrasound diagnosis [30]. In 47 (1.3%) of 3,491 participants of Framingham study according to the findings of two-dimensional echocardiography performed in compliance with the standard diagnostic criteria there was detected classic MVP (with thickening of mitral leaflets), in 37 (1.1%) nonclassic MVP with an overall prevalence of 2.4% [1]. Turker et al. [41] report much lower values of mitral prolapse prevalence. Besides, they note a fairly even distribution of the pathology among individuals in each decade from 30 to 80 years of age, and identical occurrence rate among men and women. Their findings differ from older studies [42–45] based on echocardiography in M-mode and/or observations of pedigrees that reported that MVP preferentially afflicted women and older individuals.

MVP is a pathology with genetic predisposition, which

however does not occur in newborns [21, 46], and is rarely observed in children (0.3%) [47] and young people (0.6%) [48]. These data convincingly characterize MVP as a progressing disease mainly affecting patients of middle years [1].

MVP prevalence does not depend on ethnicity. Incidence of this pathology in the population sample of American Indians (the Strong Heart Study) [2] and Canadians of South-Asian, European and Chinese origin (the SHARE study) [49] are analogous to the data presented in the Framingham study [1, 39, 40], whose participants were mainly white Americans. Similar results were obtained in the investigations based on Russian population [50].

As far as syndromic MVP in congenital disorders of the connective tissue is concerned, the following trends are noted. In Marfan syndrome, the rate of mitral valve involvement is considerable and makes about 75%, while in more severe variants with myxomatous valve alterations it approximates 28% [51]; leaflets of aortic and tricuspid valves are also subjected to the characteristic changes [52]. MVP prevalence in patients with Ehlers–Danlos syndrome is much lower (6%) [9]. A similar trend is noted in Loeys–Dietz syndrome. When 71 patients with mutations of in the *TGFBR2* gene (typical for Loeys–Dietz syndrome) and 243 patients with mutations of in the fibrillin gene (*FBN1*) (typical for Marfan syndrome) were examined, a higher MVP prevalence and mitral regurgitation were found in the last two: 45 and 56% vs. 21 and 35%, respectively [53]. Abnormalities of the mitral valve appeared also frequent among the patients with aneurysms-osteoarthritis syndrome: MVP was found in 45% of cases, mitral regurgitation in 27% [12].

### Prognosis in mitral valve prolapse

Previously, MVP was thought to be a pathology with frequent and serious complications (including stroke, atrial fibrillation, heart failure), and a high demand of surgical correction of mitral insufficiency [3, 6–8, 54, 55]. The results of the Framingham study gave grounds to consider MVP as a benign pathology with a low probability of unfavorable sequelae [1]. In the articles published at the turn of the XXI century with the eloquent titles: “Mysteries of mitral valve prolapse”, “Mitral valve prolapse: time for a fresh look”, “Mitral valve prolapse: old beliefs yield to new knowledge”, “Mitral valve prolapse: the merchant of Venice or much ado about nothing”, “When should mitral valve prolapse be considered a real disease?” the authors advocated the idea that serious complications occur in patients with the diagnosis “mitral valve prolapse” as frequent as in the individuals without it [56–60]. In some investigations, in particular, no supporting evidence was found about a close relation of MVP with a cerebral stroke [61, 62], infective endocarditis [63], and other complications [57, 63]. It may be explained by the errors of examination

methods associated with the comparison of clinically manifestant patients with practically healthy volunteers [1, 54, 55]. Reconsideration of diagnostic criteria for mitral prolapse aggravated the difference in the views on the prevalence of complications in this pathology [30]. In the Framingham study, none of the patients with MVP had a registered heart failure; the rate of atrial fibrillation, cerebral stroke, and syncope appeared to be comparable with the similar sequelae in the individuals without prolapse (1.2, 1.2, 3.6% vs. 1.7, 1.5, 3.0%) [1].

Large-scale investigations in Mayo clinic demonstrated clinical heterogeneity of MVP and various prognoses for this pathology [15]. Asymptomatic patients under 50 years of age with a normal left ventricular function have a favorable prognosis even in the presence of heavy mitral regurgitation [15, 64]. Advantages of early surgical correction of the valvular defect compared to a watchful waiting tactics in such patients remain unproved [64–67].

Mitral regurgitation is one of the main risk factors of development of unfavorable cardiovascular events in MVP (congestive heart failure, atrial fibrillation, cerebral disorders, endocarditis) as well as an indication to surgical treatment [1, 54, 55, 68–70]. In the Framingham study, asymptomatic MVP in the period from 3 to 16 years showed itself by the signs of regurgitation in a quarter of patients [71]. According to the data from Mayo clinic, the volume of mitral regurgitation increased by more than 8 ml during 1.5 years of follow-up in 51% of patients with MVP [70]. The two independent factors of the mitral regurgitation volume increase over time were progression of the valve lesion (namely, the appearance of a flail leaflet), and enlargement of a mitral annulus diameter [70]. Atrial fibrillation can also worsen the severity of mitral regurgitation but the intensity of the latter decreases after the restoration of the sinus rhythm [72–74].

In some works [54, 55] thickness of the mitral leaflet more than 5 mm (a sign of myxomatous degeneration) recorded by echocardiography in M-mode was associated with a high risk of endocarditis, mitral regurgitation, and sudden death development. Later and large-scale investigations with the application of two-dimensional echocardiography did not confirm this fact [5]. In the course of a long-term follow-up of 833 patients with asymptomatic MVP the predictors of mortality were presence of mitral regurgitation and left ventricular dysfunction at the time of primary examination, whereas age older than 50, enlargement of the left atrial cavity, and presence of mitral regurgitation were the risk factors for cardiovascular disturbances [68].

Availability of a mitral valve flail leaflet is associated with an ambiguous prognosis [75]. Asymptomatic patients with such leaflet and unaltered left ventricular function receiving medicamentous treatment have a low risk of cardiovascular complications [72, 75]. Indications for surgery on the valves in these patients are progression of atrial fibrillation (4% per year) and heart

failure (5.7% per year). Elderly age, clinical symptoms or decrease of ejection fraction by less than 60% at the time of establishing the diagnosis are markers of elevated mortality and speak in favor of performing operations on the heart valves [75, 76].

The Framingham study has demonstrated equal prevalence of MVP in men and women [1], though a community study in Olmsted County (USA) using current echocardiographic criteria showed that this pathology occurs more often in women and at a younger age than in men [15], with complications being revealed less commonly [76]. Investigation in Mayo clinic detected morphofunctional differences of heart structures in men and women with prolapse. In women, prolapse of the anterior or both mitral leaflets occurs more often, more leaflet thickening, less flail registration rate are noted [77]. Besides, they make an essential portion of patients with moderate and severe mitral regurgitation [5]. As a consequence, in a long-term perspective a higher mortality rate but an equivalent survival time after the operation on the valves are noted in women compared to men [77].

Lately, the focus of researchers' attention has shifted from the valvular mechanism to the state of the left ventricular myocardium in the assessment of prognosis in MVP. Impairment of overall hemodynamics in this pathology was shown to occur not only due mitral incompetence but via the defects of structures and functions of extracellular myocardial matrix, which can cause diastolic dysfunction, contractile capability decrease, and development of secondary cardiomyopathy [78–81].

### **Molecular biology and genetics of mitral valve prolapse**

Molecular genetic studies have significantly enriched the notion of MVP and allowed cardiologists to improve its diagnosis [8]. By the present time, participation of several genes and factors of their activation in the process of heart valves formation has been proved. Among them are calcineurin stimulating the family of nuclear factors of activated T cells (NFAT), absence of which leads to the fatal valvular defects [82]; Wnt/ $\beta$ -catenin, determining the development of endothelial cells [83]; fibroblast growth factor FGF4; homeobox gene *Sox4*; modulator of transforming growth factor beta (TGF- $\beta$ ); superfamily of signaling proteins SMAD6 [84, 85], the impaired work of which results in abnormal valve thickening. Defects in genes or signaling molecules may induce myxomatous valvular alterations and promote progressing impairment of their mechanical strength during life [86, 87].

A leading role in various mechanisms of pathogenesis of syndromic and non-syndromic MVP is played by the failure of TGF- $\beta$  regulation [21, 88]. TGF- $\beta$  is a protein controlling a number of physiological processes including angiogenesis, proliferation, cellular differentiation and

apoptosis of the majority of cells [85, 89, 90]. This representative of cytokines exerts multidirectional action on the extracellular matrix structure. Stimulation of TGF- $\beta$  via a canonic SMAD signaling pathway induces profibrotic effect including deposition of collagen and elastin [91], reduction of proteolytic enzyme expression (matrix metalloproteinases (MMP)) [92], and increase of tissue MMP inhibitor activity [93, 94]. Its stimulation via a non-canonical SMAD-independent signaling pathway results in degradation of the extracellular matrix due to the increased proteolysis via MMP-2, 9 and 13 [95], and elevated activation of MMP with the help of plasminogen activators [96].

Some works [85, 97] describe the ability of TGF- $\beta$  to initiate the development of interstitial cells of a valvular leaflet as a pathological phenotype. The importance of TGF- $\beta$  signaling pathway in emergence of sporadic MVP cases has been confirmed [98]. In the experiment on the culture of interstitial cells, induced TGF- $\beta$  production of extracellular matrix is shown to depend on SMAD2/3 and signaling protein p38 and to decelerate by angiotensin II receptor blockers [85, 98]. In the study on the surgical mitral valve specimens in patients with prolapse, it is noted that a stimulating effect of TGF- $\beta$  is secondary and occurs in response to inhibiting expression of genes regulating the response to the oxidative stress [99]. Activation of TGF- $\beta$ , in its turn, leads to the inhibition of genes responsible for proteoglycan degradation resulting in excessive accumulation of extracellular matrix [5].

Synthesis of extracellular matrix can be induced both *in vitro* and *in vivo* by mechanical stretching of the valve, which not only provides its normal development but adaptation to pathological conditions as well [86]. The ability of the valve to restore its previous parameters in response to the weakening of the mechanical action is important in the clinical context: annuloplasty by reducing the load on the valves and chord improves a long-term prognosis for patients with MVP [5].

Pathogenesis of chord rupture and generation of flails are explained by the unique properties of the extracellular matrix [72]. The major part of the valvular complex is known to be avascular just like cartilages and ligaments. A local expression of tendon-specific protein, tenomodulin, antiangiogenic properties of which have been recently discovered, is noted in the valvular chords [100]. In the areas of chord rupture tenomodulin is absent, abnormal vascular formations and intensified expression of vascular endothelial growth factor VEGF-A are observed. On the contrary, in normal unaffected chord areas, an elevated concentration of CD11b<sup>+</sup>, CD14<sup>+</sup>, and vimentin with enhanced expression of MMP-2 and MMP-13 in combination with tenomodulin inhibition is fixed [100].

Syndromic MVP related to hereditary disorders of the connective tissue manifests itself by the same myxomatous changes as primary prolapse. Marfan syndrome is associated with mutations in the *FBN1* gene located on 15q15-q21 chromosome [85, 90, 101,

102], it may also be caused by mutation in gene TGF- $\beta$  located on the 3p24.2-p25 chromosome [21, 103]. The role of *FBN1* and TGF- $\beta$  mutation in MVP pathogenesis was confirmed by the experiment on mice with the model of Marfan syndrome [104]. In the prolapsed part of the mice valves with *FBN1* deficiency, intensification of TGF- $\beta$  expression was noted. Application of antibodies neutralizing TGF- $\beta$  results in the reduction of mitral leaflet thickness and confirms the hypothesis that abnormalities of the mitral valve are linked to the increased level of TGF- $\beta$  [104]. However, a successful correction of many phenotypic manifestations of the Marfan syndrome on the mice models using TGF- $\beta$ -neutralizing antibodies is not yet applicable for treating humans [5].

Establishment of interconnection between angiotensin II and TGF- $\beta$  created premises for experimental treatment of mice with mutation in the *FBN1* gene with losartan, angiotensin II receptor antagonist [105]. It should be noted that in two experimental groups of mutant mice receiving  $\beta$ -adrenoblockers and losartan in the doses equivalent by hemodynamic effect, advantage of the latter in the ability to prevent aortic root widening was shown [105]. In the randomized study COMPARE, losartan decreased the rate of aortic root dilatation in comparison with placebo in adults with Marfan syndrome [106]. Nevertheless, the ability to reduce TGF- $\beta$  expression is noted in  $\beta$ -adrenoblockers as well [102]. In the researches of Lacro et al. [107, 108], where they compared the effect of losartan and atenolol on aorta dilatation and MVP in children and adolescents with Marfan syndrome, no essential difference between the preparations was found.

Loeys–Dietz syndrome is another disease of the connective tissue associated with the pathology of the mitral valve, which is caused by heterozygous mutations in *TGFBR1* or *TGFBR2* genes encoding subunits of TGF- $\beta$  receptors [10, 85]. In patients with this pathology, signs of TGF- $\beta$  activity are observed in the form of elevated accumulation of nuclear phosphorylated SMAD2 and concentration increase of connective tissue growth factor, which is induced by TGF- $\beta$  [10, 109]. Aneurism-osteoarthritis syndrome (combination of aortic aneurism, tortuous arteries, facial dysmorphias and early onset of osteoarthritis) caused by mutations in the *SMAD3* gene is one more confirmation of the relation of the increased expression of TGF- $\beta$  and myxomatosis of the mitral valve [12].

A high prevalence of mitral prolapse in Marfan syndrome gives grounds to suggest that primary MVP is linked with mutations in the *FBN1* gene, but nobody could prove yet this hypothesis. Unsuccessful attempts to find definite genetic defects are connected with absence of systematic study of human genome and phenotypic heterogeneity of the pathology [5].

In 1999, the first genetic locus (*MMVP1*) for non-syndromic MVP was mapped on 16p11.2-p12.1 chromosome in the family with the signs of autosomal dominant type of inheritance [110]. In 2003, the second

locus (*MMVP2*) on 11p15.4 chromosome was detected [111]. At last, in 2005, the next locus (*MMVP3*) for autosomal dominant MVP was mapped on 13q31.3-q32 chromosome [112]. Discovery of *MMVP3* confirmed genetic heterogeneity of MVP and allowed the scientists to determine the manifestation spectrum, which should be referred to hereditary pathology rather than to the variants of a norm as it has been thought previously [112].

In recent years, the so-called prodromal (for MVP development) morphology, including abnormal anterior cooptation of the leaflets, has attracted attention of investigators [71, 112]. Patients with such abnormality and minimal systolic leaflet displacement have fully or essentially the same “risk haplotype” as those having MVP, similar morphology is observed also in connection with chromosome 11 [112]. Screening of early MVP forms is necessary as rather often this pathology is clinically manifested in the fifth or sixth decade of life in the form of serious heart events [54, 55, 72, 113]. Timely reduction of hemodynamic load on the mitral valve leaflets in genetically predisposed individuals can prevent progression of MVP and appearance of severe mitral regurgitation [98, 104, 105].

A rare form of myxomatous lesion, X-linked MVP, was first described in 1969 under the name of “myxomatous valvular dystrophy” [114–116]. It is characterized by multiple myxomatous lesions of the valves, though no considerable histopathological differences from heavy forms of autosomal dominant form of MVP are observed.

The case of myxomatous valvular dystrophy combined with hemophilia A in the members of one family [115] allowed researchers to collate this variant of MVP with a sex chromosome Xq28; a combination with hemophilia considerably facilitated gene mapping. During the selection of candidate genes, P637Q missense mutation in the filamin A gene was detected in the sick members of the family [21, 115, 116]. The analysis of the mutated filamin gene in other families enabled detection of additional mutations: two new missense mutations (*G288R*, *V711D*) and deletion of bp-segment 1944 [116, 117].

Filamins are large cytoplasmic proteins providing crosslinking of actinic filament into three-dimensional structure and serving as transmitters of cellular signals [118–121]. A group of filaments is presented by A, B, and C variants, the former being responsible for the development of the heart and vessels [118, 122]. Prenatal death and a vast list of cardiovascular developmental defects including abnormal thickening and deformation of valvular leaflets are typical for hemizygous mice with a null allele of filamin A [122, 123]. Defects of other variants of filamin genes (B and C) were not accompanied by cardiovascular defects in the experiment on mutant mice [124, 125]. Pathology of the valves may be realized through the impaired signaling function of filamin A, which coordinates localization and activity of TGF- $\beta$  receptors-activators of SMAD, especially SMAD2, and can serve as a positive regulator

of TGF- $\beta$  signaling [90, 126, 127]. Mutations in the filamin A may explain the similarity of clinical manifestations of Marfan syndrome and non-syndromic MVP, since both these states are characterized by TGF- $\beta$  activity increase. In particular, myxomatosis of mitral valvular leaflets found in mice with *FBN1* deficiency reflects excessive activation of TGF- $\beta$  and impairment of filamin A expression regulation [104, 128].

Thus, at present, MVP is assumed to be the result of multiple genetic disorders, which is proved by the identification of several genes related to the occurrence of syndromic mitral prolapse [101, 103], and three loci for non-syndromic prolapse [110–112]. Detection of filamin A mutation in X-linked form of valvular dystrophy [114] confirmed the significance of cytoskeleton not only for structural integrity of the valve but also for the processes in the most important cellular signaling pathways realized, in particular, with participation of TGF- $\beta$ . Advances in the technology of DNA sequencing may lead to identification of *MMVP1*, *MMVP2*, and *MMVP3* genes in the near future. Creation of large-scale registers of patients with MVP and conduction of genome-wide studies will make it possible to identify new genes related to the occurrence of MVP, and to provide screening of asymptomatic patients for whom the development and progression of mitral regurgitation is a real threat [8, 112, 129]. New perspective ways of MVP management are feasible owing to the results of the operative material investigations *in vitro*, which demonstrated that typical myxomatous alterations can be prevented with the help of pharmacological agents [98, 102, 130]. Understanding the mechanisms of MVP development will allow the scientists to devise new methods of therapy realized via the effect on various cells and signaling pathways including inhibition of excessive TGF- $\beta$  expression by angiotensin II receptor blockers, manipulations with endothelial cells-precursors, and tissue engineering of heart valves [5, 8, 118]. Such medical approaches are especially perspective at an early stage of undiagnosed MVP phenotypes or as an alternative to surgical treatment of clinical complications in patients with a verified diagnosis.

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## References

1. Freed L.A., Levy D., Levine R.A., Larson M.G., Evans J.C., Fuller D.L., Lehman B., Benjamin E.J. Prevalence and clinical outcome of mitral-valve prolapse. *N Engl J Med* 1999; 341(1): 1–7, <http://dx.doi.org/10.1056/nejm199907013410101>.
2. Devereux R.B., Jones E.C., Roman M.J., Howard B.V., Fabsitz R.R., Liu J.E., Palmieri V., Welty T.K., Lee E.T. Prevalence and correlates of mitral valve prolapse in a population-based sample of American Indians: the Strong Heart Study. *Am J Med* 2001; 111(9): 679–685, [https://doi.org/10.1016/s0002-9343\(01\)00981-0](https://doi.org/10.1016/s0002-9343(01)00981-0).

3. Levine R.A., Slaughter S.A. Molecular genetics of mitral valve prolapse. *Curr Opin Cardiol* 2007; 22(3): 171–175, <https://doi.org/10.1097/hco.0b013e3280f3bfcf>.
4. Grau J.B., Pirelli L., Yu P.J., Galloway A.C., Ostrer H. The genetics of mitral valve prolapse. *Clin Genet* 2007; 72(4): 288–295, <https://doi.org/10.1111/j.1399-0004.2007.00865.x>.
5. Delling F.N., Vasan R.S. Epidemiology and pathophysiology of mitral valve prolapse: new insights into disease progression, genetics, and molecular basis. *Circulation* 2014; 129(21): 2158–2170, <https://doi.org/10.1161/circulationaha.113.006702>.
6. Hayek E., Gring C.N., Griffin B.P. Mitral valve prolapse. *Lancet* 2005; 365(9458): 507–518, [https://doi.org/10.1016/s0140-6736\(05\)70275-0](https://doi.org/10.1016/s0140-6736(05)70275-0).
7. Schoen F.J. Evolving concepts of cardiac valve dynamics: the continuum of development, functional structure, pathobiology, and tissue engineering. *Circulation* 2008; 118(18): 1864–1880, <https://doi.org/10.1161/circulationaha.108.805911>.
8. Levine R.A., Hagège A.A., Judge D.P., Padala M., Dal-Bianco J.P., Aikawa E., Beaudoin J., Bischoff J., Bouatia-Naji N., Bruneval P., Butcher J.T., Carpentier A., Chaput M., Chester A.H., Clusel C., Delling F.N., Dietz H.C., Dina C., Durst R., Fernandez-Friera L., Handschumacher M.D., Jensen M.O., Jeunemaitre X.P., Le Marec H., Le Tourneau T., Markwald R.R., Mérot J., Messas E., Milan D.P., Neri T., Norris R.A., Peal D., Perrocheau M., Probst V., Pucéat M., Rosenthal N., Solis J., Schott J.J., Schwammenthal E., Slaughter S.A., Song J.K., Yacoub M.H.; Leducq Mitral Transatlantic Network. Mitral valve disease — morphology and mechanisms *Nat Rev Cardiol* 2015; 12(12): 689–710, <https://doi.org/10.1038/nrcardio.2015.161>.
9. Dolan A.L., Mishra M.B., Chambers J.B., Grahame R. Clinical and echocardiographic survey of the Ehlers–Danlos syndrome. *Br J Rheumatol* 1997; 36(4): 459–462, <https://doi.org/10.1093/rheumatology/36.4.459>.
10. Loeys B.L., Schwarze U., Holm T., Callewaert B.L., Thomas G.H., Pannu H., De Backer J.F., Oswald G.L., Symoens S., Manouvrier S., Roberts A.E., Faravelli F., Greco M.A., Pyeritz R.E., Milewicz D.M., Coucke P.J., Cameron D.E., Braverman A.C., Byers P.H., De Paepe A.M., Dietz H.C. Aneurysm syndromes caused by mutations in the TGF- $\beta$  receptor. *N Engl J Med* 2006; 355(8): 788–798, <https://doi.org/10.1056/nejmoa055695>.
11. Rubegni P., Mondillo S., De Aloe G., Agricola E., Bardelli A.M., Fimiani M. Mitral valve prolapse in healthy relatives of patients with familial Pseudoxanthoma elasticum. *Am J Cardiol* 2000; 85(10): 1268–1271, [https://doi.org/10.1016/s0002-9149\(00\)00745-1](https://doi.org/10.1016/s0002-9149(00)00745-1).
12. van de Laar I.M., Oldenburg R.A., Pals G., Roos-Hesselink J.W., de Graaf B.M., Verhagen J.M., Hoedemaekers Y.M., Willemsen R., Severijnen L.A., Venselaar H., Vriend G., Pattynama P.M., Collée M., Majoorkrakauer D., Poldermans D., Frohn-Mulder I.M., Micha D., Timmermans J., Hilhorst-Hofstee Y., Bierma-Zeinstra S.M., Willems P.J., Kros J.M., Oei E.H., Oostra B.A., Wessels M.W., Bertoli-Avella A.M. Mutations in SMAD3 cause a syndromic form of aortic aneurysms and dissections with early-onset osteoarthritis. *Nat Genet* 2011; 43(2): 121–126, <https://doi.org/10.1038/ng.744>.
13. Rabkin E., Aikawa M., Stone J.R., Fukumoto Y., Libby P., Schoen F.J. Activated interstitial myofibroblasts express catabolic enzymes and mediate matrix remodeling in myxomatous heart valves. *Circulation* 2001; 104(21): 2525–2532, <https://doi.org/10.1161/hc4601.099489>.
14. Guy T.S., Hill A.C. Mitral valve prolapse. *Annu Rev Med* 2012; 63(1): 277–292, <https://doi.org/10.1146/annurev-med-022811-091602>.
15. Avierinos J.F., Gersh B.J., Melton L.J., Bailey K.R., Shub C., Nishimura R.A., Tajik A.J., Enriquez-Sarano M. Natural history of asymptomatic mitral valve prolapse in the community. *Circulation* 2002; 106(11): 1355–1361, <https://doi.org/10.1161/01.cir.0000028933.34260.09>.
16. Bayer-Topilsky T., Suri R.M., Topilsky Y., Marmor Y.N., Trenerry M.R., Antiel R.M., Mahoney D.W., Schaff H.V., Enriquez-Sarano M. Mitral valve prolapse, psychoemotional status, and quality of life: prospective investigation in the current era. *Am J Med* 2016; 129(10): 1100–1109, <https://doi.org/10.1016/j.amjmed.2016.05.004>.
17. Bonow R.O., Carabello B.A., Chatterjee K., de Leon A.C. Jr., Faxon D.P., Freed M.D., Gaasch W.H., Lytle B.W., Nishimura R.A., O’Gara P.T., O’Rourke R.A., Otto C.M., Shah P.M., Shanewise J.S. 2008 focused update incorporated into the ACC/AHA 2006 guidelines for the management of patients with valvular heart disease: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines (Writing Committee to Revise the 1998 Guidelines for the Management of Patients With Valvular Heart Disease): endorsed by the Society of Cardiovascular Anesthesiologists, Society for Cardiovascular Angiography and Interventions, and Society of Thoracic Surgeons. *Circulation* 2008; 118(15): e523–e661, <https://doi.org/10.1161/circulationaha.108.190748>.
18. Dal-Bianco J.P., Levine R.A. Anatomy of the mitral valve apparatus: role of 2D and 3D echocardiography. *Cardiol Clin* 2013; 31(2): 151–164, <https://doi.org/10.1016/j.ccl.2013.03.001>.
19. McGhie J.S., de Groot-de Laat L., Ren B., Vletter W., Frowijn R., Oei F., Geleijnse M.L. Transthoracic two-dimensional xPlane and three-dimensional echocardiographic analysis of the site of mitral valve prolapse. *Int J Cardiovasc Imaging* 2015; 31(8): 1553–1560, <https://doi.org/10.1007/s10554-015-0734-7>.
20. Loardi C., Alamanni F., Trezzi M., Kassem S., Cavallotti L., Tremoli E., Pacini D., Parolari A. Biology of mitral valve prolapse: the harvest is big, but the workers are few. *Int J Cardiol* 2011; 151(2): 129–135, <https://doi.org/10.1016/j.ijcard.2010.11.004>.
21. Padang R., Bagnall R.D., Semsarian C. Genetic basis of familial valvular heart disease. *Circ Cardiovasc Genet* 2012; 5(5): 569–580, <https://doi.org/10.1161/circgenetics.112.962894>.
22. Yagoda A.V., Novikova M.V., Gladkikh N.N. Risk factors prognostic significance of cardiac arrhythmias in connective tissue dysplasia. *Archive of internal medicine* 2015; 1: 60–63.
23. Turker Y., Ozaydin M., Acar G., Ozgul M., Hoscan Y., Varol E., Dogan A., Erdogan D., Yucel H. Predictors of ventricular arrhythmias in patients with mitral valve prolapse. *Int J Cardiovasc Imaging* 2010; 26(2): 139–145, <https://doi.org/10.1007/s10554-009-9514-6>.
24. Semyonkin A.A., Tereshchenko Yu.V., Drokina O.V., Zhivilova L.A. Autonomic regulation features in young with connective tissue dysplasia. *Sibirskiy meditsinskiy zhurnal* 2011; 26(3–2): 56–59.
25. Nedostup A.V. Some features of cardiac arrhythmia treatment in outpatient practice. *Ter Arkh* 2006; 78(8): 5–13.
26. Nechaeva G.I., Viktorova I.A., Druk I.V., Vershinina M.V.

Connective tissue dysplasia: the pulmonary aspects. *Pul'monologiya* 2004; 2: 116–119.

27. Torshin I.Yu., Gromova O.A., Kalacheva A.G., Oshchepkova E.V., Martynov A.I. Meta-analysis of clinical trials of cardiovascular effects of magnesium orotate. *Ter Arkh* 2015; 87(6): 88–97.

28. Savage D.D., Garrison R.J., Devereux R.B., Castelli W.P., Anderson S.J., Levy D., McNamara P.M., Stokes J. 3rd, Kannel W.B., Feinleib M. Mitral valve prolapse in the general population. 1. Epidemiologic features: the Framingham Study. *Am Heart J* 1983; 106(3): 571–576, [https://doi.org/10.1016/0002-8703\(83\)90704-4](https://doi.org/10.1016/0002-8703(83)90704-4).

29. Savage D.D., Devereux R.B., Garrison R.J., Castelli W.P., Anderson S.J., Levy D., Thomas H.E., Kannel W.B., Feinleib M. Mitral valve prolapse in the general population. 2. Clinical features: the Framingham Study. *Am Heart J* 1983; 106(3): 577–581, [https://doi.org/10.1016/0002-8703\(83\)90705-6](https://doi.org/10.1016/0002-8703(83)90705-6).

30. Levine R.A., Stathogiannis E., Newell J.B., Harrigan P., Weyman A.E. Reconsideration of echocardiographic standards for mitral valve prolapse: lack of association between leaflet displacement isolated to the apical four chamber view and independent echocardiographic evidence of abnormality. *J Am Coll Cardiol* 1988; 11(5): 1010–1019, [https://doi.org/10.1016/s0735-1097\(98\)90059-6](https://doi.org/10.1016/s0735-1097(98)90059-6).

31. Apor A., Nagy A.I., Kovács A., Manouras A., Andrassy P., Merkely B. Three-dimensional dynamic morphology of the mitral valve in different forms of mitral valve prolapse — potential implications for annuloplasty ring selection. *Cardiovasc Ultrasound* 2016; 14(1): 32, <https://doi.org/10.1186/s12947-016-0073-4>.

32. Russian Society of Cardiology. Hereditary disorders of connective tissue in cardiology. Diagnosis and treatment. Russian Guidelines (I revision). *Rossiyskiy kardiologicheskii zhurnal* 2013; 1(Suppl 1): 1–32.

33. Freed L.A., Benjamin E.J., Levy D., Larson M.G., Evans J.C., Fuller D.L., Lehman B., Levine R.A. Mitral valve prolapse in the general population: the benign nature of echocardiographic features in the Framingham Heart Study. *J Am Coll Cardiol* 2002; 40(7): 1298–1304, [https://doi.org/10.1016/s1062-1458\(02\)01019-x](https://doi.org/10.1016/s1062-1458(02)01019-x).

34. Maffessanti F., Mirea O., Tamborini G., Pepi M. Three-dimensional echocardiography of the mitral valve: lessons learned. *Curr Cardiol Rep* 2013; 15(7): 377, <https://doi.org/10.1007/s11886-013-0377-z>.

35. Benenstein R., Saric M. Mitral valve prolapse: role of 3D echocardiography in diagnosis. *Curr Opin Cardiol* 2012; 27(5): 465–476, <https://doi.org/10.1097/hco.0b013e328356afe9>.

36. Qamruddin S., Naqvi T.Z. Advances in 3D echocardiography for mitral valve. *Expert Rev Cardiovasc Ther* 2011; 9(11): 1431–1443, <https://doi.org/10.1586/erc.11.137>.

37. Lang R.M., Tsang W., Weinert L., Mor-Avi V., Chandra S. Valvular heart disease. The value of 3-dimensional echocardiography. *J Am Coll Cardiol* 2011; 58(19): 1933–1944, <https://doi.org/10.1016/j.jacc.2011.07.035>.

38. Jin C.N., Salgo I.S., Schneider R.J., Kam K.K., Chi W.K., So C.Y., Tang Z., Wan S., Wong R., Underwood M., Lee A.P. Using anatomic intelligence to localize mitral valve prolapse on three-dimensional echocardiography. *J Am Soc Echocardiogr* 2016; 29(10): 938–945, <https://doi.org/10.1016/j.echo.2016.07.002>.

39. Dawber T.R., Meadors G.F., Moore F.E. Jr. Epidemiological approaches to heart disease: the Framingham

Study. *Am J Public Health Nations Health* 1951; 41(3): 279–286, <https://doi.org/10.2105/ajph.41.3.279>.

40. Kannel W.B., Feinleib M., McNamara P.M., Garrison R.J., Castelli W.P. An investigation of coronary heart disease in families. The Framingham offspring study. *Am J Epidemiol* 1979; 110(3): 281–290.

41. Turker Y., Baltaci D., Basar C., Akkaya M., Ozhan H. The prevalence and clinical characteristics of mitral valve prolapse in a large population-based epidemiologic study: the MELEN study. *Eur Rev Med Pharmacol Sci* 2015; 19(12): 2208–2212.

42. Strahan N.V., Murphy E.A., Fortuin N.J., Come P.C., Humphries J.O. Inheritance of the mitral valve prolapse syndrome. Discussion of a three-dimensional penetrance model. *Am J Med* 1983; 74(6): 967–972, [https://doi.org/10.1016/0002-9343\(83\)90792-1](https://doi.org/10.1016/0002-9343(83)90792-1).

43. Zua M.S., Dziegielewski S.F. Epidemiology of symptomatic mitral valve prolapse in black patients. *J Natl Med Assoc* 1995; 87(4): 273–275.

44. Gupta R., Jain B.K., Gupta H.P., Ranawat S.S., Sharma A.K., Gupta K.D. Mitral valve prolapse: two dimensional echocardiography reveals a high prevalence in three to twelve year old children. *Indian Pediatr* 1992; 29(4): 415–423.

45. Oladapo O.O., Falase A.O. Prevalence of mitral valve prolapse in healthy adult Nigerians as diagnosed by echocardiography. *Afr J Med Med Sci* 2001; 30(1–2): 13–16.

46. Nascimento R., Freitas A., Teixeira F., Pereira D., Cardoso A., Dinis M., Mendonça I. Is mitral valve prolapse a congenital or acquired disease? *Am J Cardiol* 1997; 79(2): 226–227, [https://doi.org/10.1016/s0002-9149\(96\)00722-9](https://doi.org/10.1016/s0002-9149(96)00722-9).

47. Hickey A.J., Wilcken D.E. Age and the clinical profile of idiopathic mitral valve prolapse. *Br Heart J* 1986; 55(6): 582–586, <https://doi.org/10.1136/hrt.55.6.582>.

48. Flack J.M., Kvasnicka J.H., Gardin J.M., Gidding S.S., Manolio T.A., Jacobs D.R. Jr. Anthropometric and physiologic correlates of mitral valve prolapse in a biethnic cohort of young adults: the CARDIA study. *Am Heart J* 1999; 138(3 Pt 1): 486–492, [https://doi.org/10.1016/s0002-8703\(99\)70151-1](https://doi.org/10.1016/s0002-8703(99)70151-1).

49. Theal M., Sleik K., Anand S., Yi Q., Yusuf S., Lonn E. Prevalence of mitral valve prolapse in ethnic groups. *Can J Cardiol* 2004; 20(5): 511–515.

50. Zemtsovskiy E.V., Malev E.G. Mitral valve prolapse: a modern view of the problem. *Byulleten' Federal'nogo tsentra serdtsa, krovi i endokrinologii im. V.A. Almazova* 2011; 3: 25–30.

51. Taub C.C., Stoler J.M., Perez-Sanz T., Chu J., Isselbacher E.M., Picard M.H., Weyman A.E. Mitral valve prolapse in Marfan syndrome: an old topic revisited. *Echocardiography* 2009; 26(4): 357–364, <https://doi.org/10.1111/j.1540-8175.2008.00825.x>.

52. Gu X., He Y., Li Z., Han J., Chen J., Nixon J.V. Echocardiographic versus histologic findings in Marfan syndrome. *Tex Heart Inst J* 2015; 42(1): 30–34, <https://doi.org/10.14503/thij-13-3848>.

53. Attias D., Stheneur C., Roy C., Collod-Bérout G., Detaint D., Faivre L., Delrue M.A., Cohen L., Francannet C., Bérout C., Claustres M., Iserin F., Khau Van Kien P., Lacombe D., Le Merrer M., Lyonnet S., Odent S., Plauchu H., Rio M., Rossi A., Sidi D., Steg P.G., Ravaud P., Boileau C., Jondeau G. Comparison of clinical presentations and outcomes between patients with TGFBR2 and FBN1 mutations in Marfan syndrome and related disorders. *Circulation* 2009; 120(25): 2541–2549, <https://doi.org/10.1161/circulationaha.109.887042>.

54. Devereux R.B., Kramer-Fox R., Shear M.K., Kligfield P., Pini R., Savage D.D. Diagnosis and classification of severity of mitral valve prolapse: methodologic, biologic, and prognostic considerations. *Am Heart J* 1987; 113(5): 1265–1280, [https://doi.org/10.1016/0002-8703\(87\)90955-0](https://doi.org/10.1016/0002-8703(87)90955-0).
55. Zuppiroli A., Rinaldi M., Kramer-Fox R., Favilli S., Roman M.J., Devereux R.B. Natural history of mitral valve prolapse. *Am J Cardiol* 1995; 75(15): 1028–1032, [https://doi.org/10.1016/s0002-9149\(99\)80718-8](https://doi.org/10.1016/s0002-9149(99)80718-8).
56. Mulumudi M.S., Vivekananthan K. Mysteries of mitral valve prolapse. Proper treatment requires consideration of all clues. *Postgrad Med* 2001; 110(2): 43–44, <https://doi.org/10.3810/pgm.2001.08.994>.
57. Playford D., Weyman A.E. Mitral valve prolapse: time for a fresh look. *Rev Cardiovasc Med* 2001; 2(2): 73–81.
58. Hayek E., Griffin B. Mitral valve prolapse: old beliefs yield to new knowledge. *Cleve Clin J Med* 2002; 69(11): 889–896, <https://doi.org/10.3949/ccjm.69.11.889>.
59. Stefanadis C., Toutouzas P. Mitral valve prolapse: the merchant of Venice or much ado about nothing? *Eur Heart J* 2000; 21(4): 255–258, <https://doi.org/10.1053/ehj.1999.1926>.
60. Bensaid J. When should mitral valve prolapse be considered a real disease? *Ann Cardiol Angeiol* 2000; 49(7): 411–413.
61. Gilon D., Buonanno F.S., Joffe M.M., Leavitt M., Marshall J.E., Kistler J.P., Levine R.A. Lack of evidence of an association between mitral-valve prolapse and stroke in young patients. *N Engl J Med* 1999; 41(1): 8–13, <https://doi.org/10.1056/nejm199907013410102>.
62. Mas J.L. Cardiopathies associated with a low embolic risk. *Rev Neurol* 1999; 155(9): 677–683.
63. Koegelenberg C.F., Doubell A., Orth H., Reuter H. Infective endocarditis in the Western Cape Province of South Africa: a three-year prospective study. *QJM* 2003; 96(3): 217–225, <https://doi.org/10.1093/qjmed/hcg028>.
64. Rosenhek R., Rader F., Klaar U., Gabriel H., Krejc M., Kalbeck D., Schemper M., Maurer G., Baumgartner H. Outcome of watchful waiting in asymptomatic severe mitral regurgitation. *Circulation* 2006; 113(18): 2238–2244, <https://doi.org/10.1161/circulationaha.105.599175>.
65. Ling L.H., Enriquez-Sarano M., Seward J.B., Orszulak T.A., Schaff H.V., Bailey K.R., Tajik A.J., Frye R.L. Early surgery in patients with mitral regurgitation due to flail leaflets: a long-term outcome study. *Circulation* 1997; 96(6): 1819–1825, <https://doi.org/10.1161/01.cir.96.6.1819>.
66. Antiochos P., Muller O., Kirsch M., Agostini M., Qanadli S., Eeckhout E., Vogt P., Prêtre R., Delabays A., Jeanrenaud X., Monney P. Approach to chronic mitral regurgitation in 2016. *Rev Med Suisse* 2016; 12(520): 1042–1048.
67. Taramasso M., Gaemperli O., Maisano F. Treatment of degenerative mitral regurgitation in elderly patients. *Nat Rev Cardiol* 2015; 12(3): 177–183, <https://doi.org/10.1038/nrcardio.2014.210>.
68. Avierinos J.F., Detaint D., Messika-Zeitoun D., Mohty D., Enriquez-Sarano M. Risk, determinants, and outcome implications of progression of mitral regurgitation after diagnosis of mitral valve prolapse in a single community. *Am J Cardiol* 2008; 101(5): 662–667, <https://doi.org/10.1016/j.amjcard.2007.10.029>.
69. Enriquez-Sarano M., Avierinos J.F., Messika-Zeitoun D., Detaint D., Capps M., Nkomo V., Scott C., Schaff H.V., Tajik A.J. Quantitative determinants of the outcome of asymptomatic mitral regurgitation. *N Engl J Med* 2005; 352(9): 875–883, <https://doi.org/10.1056/nejmoa041451>.
70. Enriquez-Sarano M., Basmadjian A.J., Rossi A., Bailey K.R., Seward J.B., Tajik A.J. Progression of mitral regurgitation: a prospective Doppler echocardiographic study. *J Am Coll Cardiol* 1999; 34(4): 1137–1144, [https://doi.org/10.1016/s0735-1097\(99\)00313-7](https://doi.org/10.1016/s0735-1097(99)00313-7).
71. Delling F.N., Rong J., Larson M.G., Lehman B., Fuller D., Osypiuk E., Stantchev P., Hackman B., Manning W.J., Benjamin E.J., Levine R.A., Vasan R.S. Evolution of mitral valve prolapse: insights from the Framingham Heart Study. *Circulation* 2016; 133(17): 1688–1695, <https://doi.org/10.1161/circulationaha.115.020621>.
72. Boudoulas K.D., Boudoulas H. Floppy mitral valve (FMV)/mitral valve prolapse (MVP) and the FMV/MVP syndrome: pathophysiologic mechanisms and pathogenesis of symptoms. *Cardiology* 2013; 126(2): 69–80, <https://doi.org/10.1159/000351094>.
73. Otani K., Takeuchi M., Kaku K., Haruki N., Yoshitani H., Eto M., Tamura M., Okazaki M., Abe H., Fujino Y., Nishimura Y., Levine R.A., Otsuji Y. Evidence of a vicious cycle in mitral regurgitation with prolapse. *Circulation* 2012; 126(11 Suppl 1): S214–S221, <https://doi.org/10.1161/circulationaha.111.084178>.
74. Gertz Z.M., Raina A., Saghy L., Zado E.S., Callans D.J., Marchlinski F.E., Keane M.G., Silvestry F.E. Evidence of atrial functional mitral regurgitation due to atrial fibrillation. *J Am Coll Cardiol* 2011; 58(14): 1474–1481, <https://doi.org/10.1016/j.jacc.2011.06.032>.
75. Grigioni F., Tribouilloy C., Avierinos J.F., Barbieri A., Ferlito M., Trojette F., Tafaneli L., Branzi A., Szymanski C., Habib G., Modena M.G., Enriquez-Sarano M. Outcomes in mitral regurgitation due to flail leaflets a multicenter European study. *JACC Cardiovasc Imaging* 2008; 1(2): 133–141, <https://doi.org/10.1016/j.jcmg.2007.12.005>.
76. Ling L.H., Enriquez-Sarano M., Seward J.B., Tajik A.J., Schaff H.V., Bailey K.R., Frye R.L. Clinical outcome of mitral regurgitation due to flail leaflet. *N Engl J Med* 1996; 335(19): 1417–1423, <https://doi.org/10.1056/nejm199611073351902>.
77. Avierinos J.F., Inamo J., Grigioni F., Gersh B., Shub C., Enriquez-Sarano M. Sex differences in morphology and outcomes of mitral valve prolapse. *Ann Intern Med* 2008; 149(11): 787–795, <https://doi.org/10.7326/0003-4819-149-11-200812020-00003>.
78. De Backer J. Cardiovascular characteristics in Marfan syndrome and their relation to the genotype. *Verh K Acad Geneesk Belg* 2009; 71(6): 335–371.
79. Khan R., Sheppard R. Fibrosis in heart disease: understanding the role of transforming growth factor-beta in cardiomyopathy, valvular disease and arrhythmia. *Immunology* 2006; 118(1): 10–24, <https://doi.org/10.1111/j.1365-2567.2006.02336.x>.
80. Malev E.G., Pshepyi A.P., Vasina L.V., Reeva S.V., Timofeev E.V., Korshunova A.L. Left ventricular remodelling and diastolic dysfunction in mitral valve prolapse. *Rossiyskiy kardiologicheskij zhurnal* 2013; 100(2): 12–19.
81. Bui A.H., Roujol S., Foppa M., Kissinger K.V., Goddu B., Hauser T.H., Zimetbaum P.J., Ngo L.H., Manning W.J., Nezafat R., Delling F.N. Diffuse myocardial fibrosis in patients with mitral valve prolapse and ventricular arrhythmia. *Heart* 2017; 103(3): 204–209, <https://doi.org/10.1136/heartjnl-2016-309303>.
82. de la Pompa J.L., Timmerman L.A., Takimoto H., Yoshida H., Elia A.J., Samper E., Potter J., Wakeham A., Marengere L., Langille B.L., Crabtree G.R., Mak T.W. Role of



- the NF-ATc transcription factor in morphogenesis of cardiac valves and septum. *Nature* 1998; 392(6672): 182–186, <https://doi.org/10.1038/32419>.
83. Hurlstone A.F., Haramis A.P., Wienholds E., Begthel H., Korving J., Van Eeden F., Cuppen E., Zivkovic D., Plasterk R.H., Clevers H. The Wnt/beta-catenin pathway regulates cardiac valve formation. *Nature* 2003; 425(6958): 633–637, <https://doi.org/10.1038/nature02028>.
84. Salhiyyah K., Yacoub M.H., Chester A.H. Cellular mechanisms in mitral valve disease. *J Cardiovasc Transl Res* 2011; 4(6): 702–709, <https://doi.org/10.1007/s12265-011-9318-7>.
85. Wheeler J.B., Ikonomidis J.S., Jones J.A. Connective tissue disorders and cardiovascular complications: the indomitable role of transforming growth factor-beta signaling. *Adv Exp Med Biol* 2014; 802: 107–127, [https://doi.org/10.1007/978-94-007-7893-1\\_8](https://doi.org/10.1007/978-94-007-7893-1_8).
86. Bischoff J., Aikawa E. Progenitor cells confer plasticity to cardiac valve endothelium. *J Cardiovasc Transl Res* 2011; 4(6): 710–719, <https://doi.org/10.1007/s12265-011-9312-0>.
87. Greenhouse D.G., Murphy A., Mignatti P., Zavadil J., Galloway A.C., Balsam L.B. Mitral valve prolapse is associated with altered extracellular matrix gene expression patterns. *Gene* 2016; 586(1): 56–61, <https://doi.org/10.1016/j.gene.2016.04.004>.
88. Rizzo S., Basso C., Lazzarini E., Celeghein R., Paolin A., Gerosa G., Valente M., Thiene G., Pilichou K. TGF-beta1 pathway activation and adherens junction molecular pattern in nonsyndromic mitral valve prolapse. *Cardiovasc Pathol* 2015; 24(6): 359–367, <https://doi.org/10.1016/j.carpath.2015.07.009>.
89. Bertolino P., Deckers M., Lebrin F., ten Dijke P. Transforming growth factor-beta signal transduction in angiogenesis and vascular disorders. *Chest* 2005; 128(6 Suppl): 585S–590S, [https://doi.org/10.1378/chest.128.6\\_suppl.585s](https://doi.org/10.1378/chest.128.6_suppl.585s).
90. LaHaye S., Lincoln J., Garg V. Genetics of valvular heart disease. *Curr Cardiol Rep* 2014; 16(6): 487, <https://doi.org/10.1007/s11886-014-0487-2>.
91. Ignatz R.A., Massague J. Transforming growth factor-beta stimulates the expression of fibronectin and collagen and their incorporation into the extracellular matrix. *J Biol Chem* 1986; 261(9): 4337–4345.
92. Yan C., Boyd D.D. Regulation of matrix metalloproteinase gene expression. *J Cell Physiol* 2007; 211(1): 19–26, <https://doi.org/10.1002/jcp.20948>.
93. Kwak H.J., Park M.J., Cho H., Park C.M., Moon S.I., Lee H.C., Park I.C., Kim M.S., Rhee C.H., Hong S.I. Transforming growth factor-beta1 induces tissue inhibitor of metalloproteinase-1 expression via activation of extracellular signal-regulated kinase and Sp1 in human fibrosarcoma cells. *Mol Cancer Res* 2006; 4(3): 209–220, <https://doi.org/10.1158/1541-7786.mcr-05-0140>.
94. Jones J.A., Spinale F.G., Ikonomidis J.S. Transforming growth factor-beta signaling in thoracic aortic aneurysm development: a paradox in pathogenesis. *J Vasc Res* 2009; 46(2): 119–137, <https://doi.org/10.1159/000151766>.
95. Kim E.S., Kim M.S., Moon A. TGF-beta-induced upregulation of MMP-2 and MMP-9 depends on p38 MAPK, but not ERK signaling in MCF10A human breast epithelial cells. *Int J Oncol* 2004; 25(5): 1375–1382, <https://doi.org/10.3892/ijco.25.5.1375>.
96. Laiho M., Saksela O., Keski-Oja J. Transforming growth factor beta alters plasminogen activator activity in human skin fibroblasts. *Exp Cell Res* 1986; 164(2): 399–407, [https://doi.org/10.1016/0014-4827\(86\)90038-8](https://doi.org/10.1016/0014-4827(86)90038-8).
97. Disatian S., Ehrhart E.J. 3rd, Zimmerman S., Orton E.C. Interstitial cells from dogs with naturally occurring myxomatous mitral valve disease undergo phenotype transformation. *J Heart Valve Dis* 2008; 17(4): 402–411.
98. Geirsson A., Singh M., Ali R., Abbas H., Li W., Sanchez J.A., Hashim S., Tellides G. Modulation of transforming growth factor-beta signaling and extracellular matrix production in myxomatous mitral valves by angiotensin II receptor blockers. *Circulation* 2012; 126(11 Suppl 1): 189–197, <https://doi.org/10.1161/circulationaha.111.082610>.
99. Hulin A., Deroanne C., Lambert C., Defraigne J.O., Nusgens B., Radermecker M., Colige A. Emerging pathogenic mechanisms in human myxomatous mitral valve: lessons from past and novel data. *Cardiovasc Pathol* 2013; 22(4): 245–250, <https://doi.org/10.1016/j.carpath.2012.11.001>.
100. Kimura N., Shukunami C., Hakuno D., Yoshioka M., Miura S., Docheva D., Kimura T., Okada Y., Matsumura G., Shin'oka T., Yozu R., Kobayashi J., Ishibashi-Ueda H., Hiraki Y., Fukuda K. Local tenomodulin absence, angiogenesis, and matrix metalloproteinase activation are associated with the rupture of the chordae tendineae cordis. *Circulation* 2008; 118(17): 1737–1747, <https://doi.org/10.1161/circulationaha.108.780031>.
101. Dietz H.C., Cutting G.R., Pyeritz R.E., Maslen C.L., Sakai L.Y., Corson G.M., Puffenberger E.G., Hamosh A., Nanthakumar E.J., Curristin S.M., Stetten G., Meyers D.A., Francomano C.A. Marfan syndrome caused by a recurrent de novo missense mutation in the fibrillin gene. *Nature* 1991; 352(6333): 337–339, <https://doi.org/10.1038/352337a0>.
102. Kumar A., Agarwal S. Marfan syndrome: an eyesight of syndrome. *Meta Gene* 2014; 2: 96–105, <https://doi.org/10.1016/j.mgene.2013.10.008>.
103. Mizuguchi T., Collod-Beroud G., Akiyama T., Abifadel M., Harada N., Morisaki T., Allard D., Varret M., Claustres M., Morisaki H., Ihara M., Kinoshita A., Yoshiura K., Junien C., Kajii T., Jondeau G., Ohta T., Kishino T., Furukawa Y., Nakamura Y., Niikawa N., Boileau C., Matsumoto N. Heterozygous TGFBR2 mutations in Marfan syndrome. *Nat Genet* 2004; 36(8): 855–860, <https://doi.org/10.1038/ng1392>.
104. Ng C.M., Cheng A., Myers L.A., Martinez-Murillo F., Jie C., Bedja D., Gabrielson K.L., Hausladen J.M., Mecham R.P., Judge D.P., Dietz H.C. TGF-beta-dependent pathogenesis of mitral valve prolapse in a mouse model of Marfan syndrome. *J Clin Invest* 2004; 114(11): 1586–1592, <https://doi.org/10.1172/jci22715>.
105. Habashi J.P., Doyle J.J., Holm T.M., Aziz H., Schoenhoff F., Bedja D., Chen Y., Modiri A.N., Judge D.P., Dietz H.C. Angiotensin II type 2 receptor signaling attenuates aortic aneurysm in mice through ERK antagonism. *Science* 2011; 332(6027): 361–365, <https://doi.org/10.1126/science.1192152>.
106. Groenink M., den Hartog A.W., Franken R., Radonic T., de Waard V., Timmermans J., Scholte A.J., van den Berg M.P., Spijkerboer A.M., Marquering H.A., Zwinderman A.H., Mulder B.J. Losartan reduces aortic dilatation rate in adults with Marfan syndrome: a randomized controlled trial. *Eur Heart J* 2013; 34(45): 3491–500, <https://doi.org/10.1093/eurheartj/eh334>.
107. Lacro R.V., Guey L.T., Dietz H.C., Pearson G.D., Yetman A.T., Gelb B.D., Loeys B.L., Benson D.W., Bradley T.J., De Backer J., Forbus G.A., Klein G.L., Lai W.W., Levine J.C., Lewin M.B., Markham L.W., Paridon S.M., Pierpont M.E., Radojewski E., Selamet Tierney E.S.,

Sharkey A.M., Wechsler S.B., Mahony L.; Pediatric Heart Network Investigators. Characteristics of children and young adults with Marfan syndrome and aortic root dilation in a randomized trial comparing atenolol and losartan therapy. *Am Heart J* 2013; 165(5): 828–835.e3, <https://doi.org/10.1016/j.ahj.2013.02.019>.

**108.** Lacro R.V., Dietz H.C., Sleeper L.A., Yetman A.T., Bradley T.J., Colan S.D., Pearson G.D., Selamet Tierney E.S., Levine J.C., Atz A.M., Benson D.W., Braverman A.C., Chen S., De Backer J., Gelb B.D., Grossfeld P.D., Klein G.L., Lai W.W., Liou A., Loeys B.L. Atenolol versus losartan in children and young adults with Marfan's syndrome. *N Engl J Med* 2014; 371(22): 2061–2071, <https://doi.org/10.1056/nejmoa1404731>.

**109.** Judge D.P., Rouf R., Habashi J., Dietz H.C. Mitral valve disease in Marfan syndrome and related disorders. *J Cardiovasc Transl Res* 2011; 4(6): 741–747, <https://doi.org/10.1007/s12265-011-9314-y>.

**110.** Disse S., Abergel E., Berrebi A., Houot A.M., Le Heuzey J.Y., Diebold B., Guize L., Carpentier A., Corvol P., Jeunemaitre X. Mapping of a first locus for autosomal dominant myxomatous mitral-valve prolapse to chromosome 16p11.2-p12.1. *Am J Hum Genet* 1999; 65(5): 1242–1251, <https://doi.org/10.1086/302624>.

**111.** Freed L.A., Acierno J.S., Dai D., Leyne M., Marshall J.E., Nesta F., Levine R.A., Slaugenhaupt S.A. A locus for autosomal dominant mitral valve prolapse on chromosome 11p15.4. *Am J Hum Genet* 2003; 72(6): 1551–1559, <https://doi.org/10.1086/375452>.

**112.** Nesta F., Leyne M., Yosefy C., Simpson C., Dai D., Marshall J.E., Hung J., Slaugenhaupt S.A., Levine R.A. New locus for autosomal dominant mitral valve prolapse on chromosome 13: clinical insights from genetic studies. *Circulation* 2005; 112(13): 2022–2030, <https://doi.org/10.1161/circulationaha.104.516930>.

**113.** Klemenov A.V. Idiopathic mitral valve prolapse in adulthood and old age. *Klinicheskaya gerontologiya* 2001; 7(5–6): 57–59.

**114.** Tourneau T., Lardeux A., Kyndt F., Mérot J., Hagege A., Levine R., Marec H., Schott J.-J., Probst V. New findings in mitral valve prolapse related to filamin-A mutations. *Archives of Cardiovascular Diseases Supplements* 2012; 4(1): 59, [https://doi.org/10.1016/s1878-6480\(12\)70583-9](https://doi.org/10.1016/s1878-6480(12)70583-9).

**115.** Kyndt F., Schott J.J., Trochu J.N., Baranger F., Herbert O., Scott V., Fressinaud E., David A., Moisan J.P., Bouhour J.B., Le Marec H., Benichou B. Mapping of X-linked myxomatous valvular dystrophy to chromosome Xq28. *Am J Hum Genet* 1998; 62(3): 627–632, <https://doi.org/10.1086/301747>.

**116.** Lardeux A., Kyndt F., Lecointe S., Marec H., Mérot J., Schott J.J., Tourneau T., Probst V. Filamin-A-related myxomatous mitral valve dystrophy: genetic, echocardiographic and functional aspects. *J Cardiovasc Transl Res* 2011; 4(6): 748–756, <https://doi.org/10.1007/s12265-011-9308-9>.

**117.** Duval D., Labbé P., Bureau L., Le Tourneau T., Norris R.A., Markwald R.R., Levine R., Schott J.J., Mérot J. MVP-associated filamin A mutations affect FlnA-PTPN12(PTP-PEST) interactions. *J Cardiovasc Dev Dis* 2015; 2(3): 233–247, <https://doi.org/10.3390/jcdd2030233>.

**118.** Nakamura F., Stossel T.P., Hartwig J.H. The filamins: organizers of cell structure and function. *Cell Adh Migr* 2011; 5(2): 160–169, <https://doi.org/10.4161/cam.5.2.14401>.

**119.** Ciobanasu C., Faivre B., Le Clainche C. Integrating

actin dynamics, mechanotransduction and integrin activation: the multiple functions of actin binding proteins in focal adhesions. *Eur J Cell Biol* 2013; 92(10–11): 339–348, <https://doi.org/10.1016/j.ejcb.2013.10.009>.

**120.** Razinia Z., Mäkelä T., Ylännä J., Calderwood D.A. Filamins in mechanosensing and signaling. *Annu Rev Biophys* 2012; 41: 227–246, <https://doi.org/10.1146/annurev-biophys-050511-102252>.

**121.** Jahed Z., Shams H., Mehrbod M., Mofrad M.R. Mechanotransduction pathways linking the extracellular matrix to the nucleus. *Int Rev Cell Mol Biol* 2014; 310: 171–220, <https://doi.org/10.1016/b978-0-12-800180-6.00005-0>.

**122.** Norris R.A., Moreno-Rodriguez R., Wessels A., Merot J., Bruneval P., Chester A.H., Yacoub M.H., Hagege A., Slaugenhaupt S.A., Aikawa E., Schott J.J., Lardeux A., Harris B.S., Williams L.K., Richards A., Levine R.A., Markwald R.R. Expression of the familial cardiac valvular dystrophy gene, filamin-A, during heart morphogenesis. *Dev Dyn* 2010; 239(7): 2118–2127, <https://doi.org/10.1002/dvdy.22346>.

**123.** Sauls K., Toomer K., Williams K., Johnson A.J., Markwald R.R., Hajdu Z., Norris R.A. Increased infiltration of extra-cardiac cells in myxomatous valve disease. *J Cardiovasc Dev Dis* 2015; 2(3): 200–213, <https://doi.org/10.3390/jcdd2030200>.

**124.** Dalkilic I., Schienda J., Thompson T.G., Kunkel L.M. Loss of filamin C (FLNc) results in severe defects in myogenesis and myotube structure. *Mol Cell Biol* 2006; 26(17): 6522–6534, <https://doi.org/10.1128/mcb.00243-06>.

**125.** Zhou X., Tian F., Sandzén J., Cao R., Flaberg E., Szekely L., Cao Y., Ohlsson C., Bergo M.O., Borén J., Akyürek L.M. Filamin B deficiency in mice results in skeletal malformations and impaired microvascular development. *Proc Natl Acad Sci USA* 2007; 104(10): 3919–3924, <https://doi.org/10.1073/pnas.0608360104>.

**126.** Sasaki A., Masuda Y., Ohta Y., Ikeda K., Watanabe K. Filamin associates with Smads and regulates transforming growth factor-beta signaling. *J Biol Chem* 2001; 276(21): 17871–17877, <https://doi.org/10.1074/jbc.m008422200>.

**127.** Cushing M.C., Liao J.T., Anseth K.S. Activation of valvular interstitial cells is mediated by transforming growth factor-beta1 interactions with matrix molecules. *Matrix Biol* 2005; 24(6): 428–437, <https://doi.org/10.1016/j.matbio.2005.06.007>.

**128.** Charitakis K., Basson C.T. Degenerating heart valves: fill them up with filamin? *Circulation* 2006; 115(1): 2–4, <https://doi.org/10.1161/circulationaha.106.663237>.

**129.** Dina C., Bouatia-Naji N., Tucker N., Delling F.N., Toomer K., Durst R., Perrocheau M., Fernandez-Friera L., Solis J., Le Tourneau T., Chen M.H., Probst V., Bosse Y., Pibarot P., Zelenika D., Lathrop M., Hercberg S., Roussel R., Benjamin E.J., Bonnet F., Lo S.H., Dolmatova E., Simonet F., Lecointe S., Kyndt F., Redon R., Le Marec H., Froguel P., Ellinor P.T., Vasan R.S., Bruneval P., Markwald R.R., Norris R.A., Milan D.J., Slaugenhaupt S.A., Levine R.A., Schott J.J., Hagege A.A., Jeunemaitre X. Genetic association analyses highlight biological pathways underlying mitral valve prolapse. *Nat Genet* 2015; 47(10): 1206–1211, <https://doi.org/10.1038/ng.3383>.

**130.** Siordia J.A. Current discoveries and interventions for barlow's disease. *Curr Cardiol Rep* 2016; 18(8): 73, <https://doi.org/10.1007/s11886-016-0754-5>.